

Statistical and Fractal Processing of Phase Images of Human Biological Fluids

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Abstract—Performed in this work are complex statistical and fractal analyses of phase properties inherent to birefringence networks of liquid crystals consisting of optically-thin layers prepared from human bile. Within the framework of a statistical approach, the authors have investigated values and ranges for changes of statistical moments of the 1-st to 4-th orders that characterize coordinate distributions for phase shifts between orthogonal components of amplitudes inherent to laser radiation transformed by human bile with various pathologies. Using the Gramm-Charlie method, ascertained are correlation criteria for differentiation of phase maps describing pathologically changed liquid-crystal networks. In the framework of the fractal approach, determined are dimensionalities of self-similar coordinate phase distributions as well as features of transformation of logarithmic dependences for power spectra of these distributions for various types of human pathologies.

Index Terms—birefringence, biological fluid, correlation, Jones Matrix, liquid crystal, phase, polarization.

I. INTRODUCTION

Among the methods for optical diagnostics of human biological tissues (BT), the methods of laser polarimetric diagnostics aimed at their optically-anisotropic structure are widely spread [1 - 31]. The main “information product” of these methods is availability of coordinate distributions for azimuths $\alpha(x, y)$ and ellipticity $\beta(x, y)$ of polarization (polarization maps) with the following types of analyses: statistical (statistical moments of the 1-st to 4-th orders [5, 6, 10, 14, 19, 25, 26, 30]), correlation (auto- and joint correlation function [12, 17, 18, 21, 26]), fractal (fractal dimensionalities [5, 6, 25]), singular (distributions of amounts of linearly and circularly polarized states [22, 28]). As a result, interrelations between the set of these parameters and distributions of optical axis directions as well as values of the birefringence characterizing the network of optically uniaxial protein (myosin, collagen, elastin, etc.) fibrils in optically anisotropic components of BT layers can be determined. Using this base, developed is a set of methods for early recognition and differentiation of pathological changes in BT structures related with their degenerative-dystrophic and oncological changes [4-6, 12, 19, 20-22, 27, 29, 31].

It is noteworthy that there exists a widespread group of optically anisotropic biological objects for which the methods of laser polarimetric diagnostics are not so efficient. Optically-thin (coefficient of extinction $\tau \leq 0.1$) layers of various biological liquids (bile, urine, liquor,

synovial liquid, blood plasma, etc.) can be related to these objects. All these layers possess considerably less optical anisotropy of the biological component matter as compared with birefringent BT structures [4]. As a consequence, these objects weakly modulate polarization of laser radiation

($\begin{cases} \alpha(x, y) \approx const; \\ \beta(x, y) \rightarrow 0. \end{cases}$). On the other hand, the biological liquids

are more available for a direct laboratory analysis as compared to traumatic methods of BT biopsy. From the above reasoning, it seems topical to search new, additional parameters for laser diagnostics of optically anisotropic structures in biological liquids.

Our work is aimed at searching the possibilities to perform diagnostics of structures inherent to liquid-crystal networks of human bile with various pathologies by using the method to determine the coordinate distributions of phase shifts (phase maps) between orthogonal components of laser radiation amplitudes with the following statistical, correlation and fractal analyses of these distributions.

II. OPTICAL MODEL OF HUMAN BILE

As a base for modeling the optical properties of human bile we use the conception of anisotropy observed in BT protein networks developed in [1-4, 7, 9, 14, 16, 23-27, 30]:

human bile can be considered as a two-component amorphous-crystalline structure;

optically isotropic - optically homogeneous micellar solution;

optically anisotropic - liquid-crystalline phase, consisting of three types of liquid crystals: needle crystals of fatty acids, cholesterol monohydrate crystals, bilirubinate crystals of calcium.

The optical properties of amorphous $\{A\}$ and crystalline $\{C\}$ components of biological liquids can be exhaustively described using the following Jones operators [26]

$$\{A\} = \begin{vmatrix} a_{11} & a_{12} \\ a_{21} & a_{22} \end{vmatrix} = \begin{vmatrix} \exp(-\tau) & 0 \\ 0 & \exp(-\tau) \end{vmatrix}; \quad (1)$$

$$\{C\} = \begin{vmatrix} c_{11} & c_{12} \\ c_{21} & c_{22} \end{vmatrix} = \begin{vmatrix} \cos^2 \rho + \sin^2 \rho \exp(-i\delta) & \cos \rho \sin \rho [1 - \exp(-i\delta)] \\ \cos \rho \sin \rho [1 - \exp(-i\delta)] & \sin^2 \rho + \cos^2 \rho \exp(-i\delta) \end{vmatrix}; \quad (2)$$

Here, τ is the absorption coefficient for laser radiation in the biological liquid layer with the geometric thickness l ; ρ - direction of the optical axis; $\delta = 2\pi/\lambda \Delta n d$ - phase shift between the orthogonal components E_x and E_y of the amplitude of illuminating laser light with the wavelength λ ; Δn - index of birefringence.

The Jones matrix of the biological liquid layer, where isotropic and anisotropic creations lie in one plane, can be expressed as a sum of operators $\{A\}$ and $\{C\}$

$$\{M\} = \{A\} + \{C\} = \begin{pmatrix} a_{11} + c_{11} & a_{12} + c_{12} \\ a_{21} + c_{21} & a_{22} + c_{22} \end{pmatrix}, \quad (3)$$

Let us consider the process of transformation of the complex amplitude ($E \rightarrow U$) of a laser wave that passed through the biological liquid layer ($\{M\}$) located between two crossed phase filters – quarter-wave plates ($\{\Phi_1\}$ and $\{\Phi_2\}$) and polarizers ($\{P_1\}$ and $\{P_2\}$), planes of transmission for which make $+45^\circ$ and -45° angles with axes of the highest velocity. The amplitude U of the transformed laser beam in this experimental setup can be determined from the following matrix equation

$$U = 0.25 \{P_2\} \{\Phi_2\} \{M\} \{\Phi_1\} \{P_1\} E. \quad (4)$$

Here,

$$\left\{ \begin{array}{l} E = \begin{pmatrix} E_x \\ E_y \exp(-i\delta_0) \end{pmatrix}, \quad U = \begin{pmatrix} U_x \\ U_y \exp(-i\delta) \end{pmatrix}, \\ \{P_1\} = \begin{pmatrix} 1 & 1 \\ 1 & 1 \end{pmatrix}, \quad \{P_2\} = \begin{pmatrix} 1 & -1 \\ -1 & 1 \end{pmatrix}, \\ \{\Phi_1\} = \begin{pmatrix} 1 & 0 \\ 0 & i \end{pmatrix}, \quad \{\Phi_2\} = \begin{pmatrix} i & 0 \\ 0 & 1 \end{pmatrix}. \end{array} \right. \quad (5)$$

In the special case of a plane-polarized wave

$$E(E_x = E_y; \delta_0 = 0) = \begin{pmatrix} 1 \\ 1 \end{pmatrix}, \text{ Eq. (4) acquires the look}$$

$$U = 0.25 \begin{pmatrix} 1 & -1 \\ -1 & 1 \end{pmatrix} \begin{pmatrix} i & 0 \\ 0 & 1 \end{pmatrix} \times \begin{pmatrix} \cos^2 \rho + \sin^2 \rho \exp[-i\delta] & \cos \rho \sin \rho [1 - \exp[-i\delta]] \\ \cos \rho \sin \rho [1 - \exp[-i\delta]] & \sin^2 \rho + \cos^2 \rho \exp[-i\delta] \end{pmatrix} \times \begin{pmatrix} 1 & 0 \\ 0 & i \end{pmatrix} \begin{pmatrix} 1 \\ 1 \end{pmatrix} \quad (6)$$

The solution of the matrix equation (6) is the value of complex amplitude $U(\delta)$ that is determined exclusively by the phase shift δ and does not depend on orientation of the optical axis ρ for a laser image of biological liquid. Being based on it, one can write

$$I(\delta) = UU^* = I_0 \sin^2 \left[\frac{\delta}{2} \right]. \quad (7)$$

Here, I_0 is the intensity of a probing laser beam, $I_\delta(r)$ - intensity of the laser image for the biological liquid layer in the point (r).

Interrelations (4) to (7) define the algorithm for direct experimental measuring the coordinate distribution of phase shifts $\delta(r)$ between orthogonal components of the amplitudes U_x, U_y in the laser image of an optically anisotropic biological liquid layer.

III. OPTICAL SCHEME AND METHOD FOR EXPERIMENTAL MEASURING THE PHASE MAPS OF BIOLOGICAL LIQUIDS

Shown in figure 1 is the optical scheme for phasometry of laser images obtained for biological liquids [4].

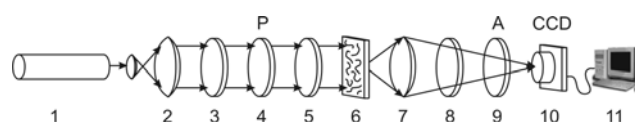


Figure 1. Optical scheme of the phase meter. 1 – He-Ne laser; 2 – collimator; 3, 5, 8 – quarter-wave plates; 4, 9 – polarizer and analyzer, respectively; 6 – investigated object; 7 – micro-objective; 10 – CCD camera; 11 – personal computer

Illumination was carried out using the parallel beam ($\varnothing = 10^4 \mu\text{m}$) of He-Ne laser 1 ($\lambda = 0.6328 \mu\text{m}$). Using the polarization illuminator (quarter-wave plate 3 and polarizer 4), we formed the beam linearly polarized with the azimuth 45° . The axis of the highest velocity of the quarter-wave plate 5 was oriented at the angle $\Theta = 45^\circ$ relatively to the transmission plane of the polarizer 4. Images of biological liquid layers 6 were projected using the micro-objective 7 into the plane of the light-sensitive area ($m \times n = 800 \text{ pix} \times 600 \text{ pix}$) of a CCD camera 10. Rotating the transmission plane of the analyzer 9 by the angle $\Theta = -45^\circ$ relatively to the axis of the highest velocity of the quarter-wave plate 8, formed were the conditions for transmission of left-circular polarized oscillations of points in laser images of biological liquids. The intensity of these oscillations I_δ was registered by each separate pixel in the CCD camera 10. Thus, we obtained the two-dimensional

$$\text{distribution } I_\delta \begin{pmatrix} r_{11} & \dots & r_{1n} \\ \dots & \dots & \dots \\ r_{m1} & \dots & r_{mn} \end{pmatrix} \text{ for this intensity. Further, in}$$

accord with (7), we calculated coordinate distributions (phase maps) for phase shifts $\delta(m \times n)$ between orthogonal components of the amplitude for laser images of biological liquids.

IV. ALGORITHMS FOR STATISTICAL, CORRELATION AND FRACTAL ANALYSES OF PHASE MAPS

To estimate $\delta(m \times n)$ distributions, we calculated the set of statistical moments of the 1-st to 4-th orders $M_{j=1,2,3,4}$ [5, 6, 25, 30]

$$M_1 = \frac{1}{N} \sum_{i=1}^N |\delta_i|, \quad M_2 = \sqrt{\frac{1}{N} \sum_{i=1}^N \delta_i^2}, \quad (8)$$

$$M_3 = \frac{1}{M_2^3} \frac{1}{N} \sum_{i=1}^N \delta_i^3, \quad M_4 = \frac{1}{M_2^4} \frac{1}{N} \sum_{i=1}^N \delta_i^4.$$

As a base for the analysis of the coordinate structure of $\delta(m \times n)$, we took the autocorrelation method using the function [12, 21, 26]

$$K_{i=1 \div n}(\Delta m) = \lim_{m \rightarrow 0} \frac{1}{m} \int_1^m [\delta_i(m)] [\delta_i(m - \Delta m)] dm. \quad (9)$$

Here, $(\Delta m = 1 \text{ pix})$ is the step of changing the coordinate $(x = 1 \div m)$ of a distribution in the set of polarization distributions $\delta(m)$ for a specific i -th line. The overall expression for the autocorrelation function is obtained by averaging the expression (9) over all the lines $i = 1 \div n$

$$K(\Delta m) = \frac{\sum_{i=1}^n K_i(\Delta m)}{n}. \quad (10)$$

As correlation parameters that characterize the dependences $K(\Delta m)$, we used:

correlation area S

$$S = \int_1^m K(\Delta m) dm, \quad (11)$$

normalized fourth statistical moment Q_4 that determine the excess of the autocorrelation function $K(\Delta m)$

$$Q_4 = \frac{N}{\left(\sum_{i=1}^N (K(\Delta m))_i^2 \right)^2} \sum_{i=1}^N (K(\Delta m))_i^4; \quad (12)$$

The fractal analysis of $\delta(m \times n)$ distributions was performed using the calculation of logarithmic dependences $\log J(\delta) - \log d^{-1}$ for the power spectra $J(\delta)$

$$J(\delta) = \int_{-\infty}^{+\infty} \delta \cos 2\pi \nu d \nu, \quad (13)$$

where $\nu = d^{-1}$ are spatial frequencies that are determined by geometrical dimensions (d) inherent to structural

elements of laser images for biological liquids.

The dependences $\log J(\delta) - \log d^{-1}$ are approximated using the least-squares method into the curves $V(\eta)$, straight parts of which are used to determine the slope angles η and fractal dimensionalities F in accord with the relation [5, 6, 11, 25]

$$F = 3 - \text{tg } \eta. \quad (14)$$

Classification of coordinate $\delta(m \times n)$ distributions is fulfilled using the following criteria:

they are fractal with the proviso that the slope angle is constant ($\eta = \text{const}$) for 2 – 3 decades of changing the sizes d ;

they are multi-fractal on the condition that there available are several slope angles $V(\eta)$;

they are random, if any stable slope angles $V(\eta)$ do not exist within the whole range of changing the sizes d .

In the latter case, the $\log J(\delta) - \log d^{-1}$ distributions are characterized with the dispersion

$$D = \sqrt{\frac{1}{N} \sum_{i=1}^N [\log J(\delta) - \log d^{-1}]^2} \quad (15)$$

V. STATISTICAL, CORRELATION AND FRACTAL PARAMETERS OF PHASE MAPS FOR HUMAN BILE

As objects for experimental studying, we chose optically-thin layers of synovial liquid taken from a healthy patient (figure 2a) and with atrophic arthritis (figure 2b).

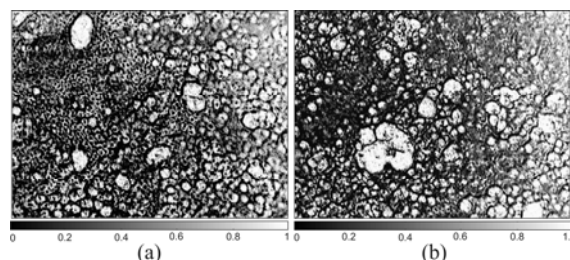


Figure 2. Polycrystalline networks of human bile taken from healthy patient (a) and patient suffering from diabetes of 2nd level (b).

The images of layers prepared from human bile (figure 2) are indicative of availability of two fractions – optically isotropic and liquid-crystal network (anisotropic one). As seen, geometric structure and sizes of separate elements in the polycrystalline network of the samples prepared from biological liquids are individual for physiological state.

Shown in figure 3 are the phase maps (fragments (a), (b), (c), (d)) and histograms (fragments (e), (f)) for distributions of random values inherent to the phase shifts δ between orthogonal components of the laser radiation amplitude transformed inside layers of synovial liquid taken from healthy patient's (left column) and from patient suffering from diabetes of 2nd level (right column).

The obtained data show that the value of phase shifts δ for laser radiation transformed inside layers of human bile lies within the short range of changes $0 \leq \delta \leq \pi$. The weak

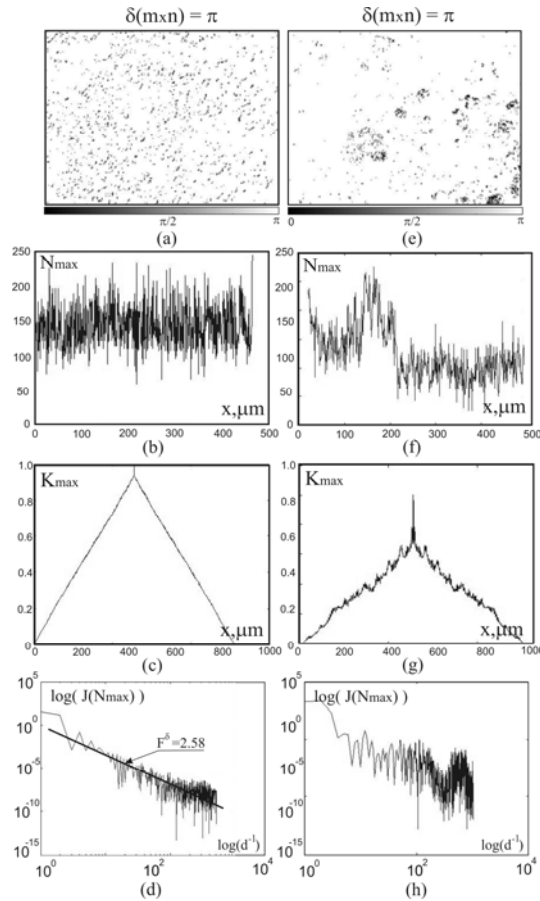


Figure 5. Coordinate $(m \times n)$ (a, e), quantitative $N_{\max}(x)$ (b, f), correlation $K_{\max}(\Delta x)$ (c, g) and fractal $\log J(N_{\max}) - \log d^{-1}$ (d, h) parameters of the extreme sample $\delta(m \times n) = 1$ for phase maps of the samples of bile belonging to a healthy patient (a, b, c, d) and a patient with diabetes of 2nd level (e, f, g, h).

The comparative analysis of the obtained set of experimental data about statistical, correlation and fractal structures in dependences for the amount of extreme values $N_{\min, \max}(x)$ inherent to phase maps describing layers of bile of healthy patient and that sick with diabetes of 2nd level enabled to found:

- tendency to a decreasing (increasing) total amount of extreme values $\delta_{\min} \rightarrow 0$ ($\delta_{\max} \rightarrow \pi$) of the phase shifts in laser images of layers prepared from bile of a patient with diabetes of 2nd level (figures 4 and 5, fragments (b, f));
- fact that autocorrelation functions $K_{\min}(\Delta x)$ (figure 4, fragments (c, g)) monotonically drop with increasing the step of scanning Δx in dependences $N_{\min}(x)$;
- correlation structure of the distribution for the extreme sample $\delta(m \times n) = \pi$ in the phase map describing the polycrystalline component in bile of a sick patient changes: at the background of monotonic drop there arise oscillations of values in the dependence $K_{\max}(\Delta x)$ (see figure 5, fragment (g));
- logarithmic dependences for the power spectra of distributions $N_{\min}(x)$ for the optically isotropic component in bile of both types possess a stable slope angle (figure 4, fragments (d, f)) within the whole range of geometric sizes inherent to the laser image registered by the CCD camera (figure 1);

- fractal distributions $N_{\max}(x)$ for phase maps of laser images describing the optically anisotropic fraction of bile a healthy man (figure 5, fragment (d)) are transformed into the statistical ones in the case of diabetes of 2nd level: approximating curve in the dependence $\log J(N_{\max}) - \log d^{-1}$ has no stable slope (Fig. 5, fragment (h)).

From the quantitative viewpoint, the dependences $N_{\min, \max}(x)$ illustrate statistical $M_{i=1-4}^{\delta}$, correlation $R^{\delta}, S^{\delta}, Q^{\delta}$ and fractal F^{δ}, D^{δ} parameters determined within the limits of two patient groups, and they are summarized in Tables 2.

Our analysis of the parameters determined experimentally has shown that the following parameters are diagnostically sensitive in observation of inflammatory processes

- statistical moments of the third (M_3^{δ}) and fourth (M_4^{δ}) orders in distributions for the amount of extreme values $N_{\max}(x)$ of phase shifts $\delta(m \times n) = 1$ in laser images for bile of both types – differences between them reach 2.4 and 4.1 times;
- excess (R_4^{δ}) of autocorrelation functions $K_{\max}(\Delta x)$ related to distributions $N_{\max}(x)$ differ by 1.8 and 2.7 times;

TABLE 2. STATISTICAL MOMENTS $M_{i=1-4}^{\delta}$, CORRELATION $R^{\delta}, S^{\delta}, Q^{\delta}$ AND FRACTAL F^{δ}, D^{δ} PARAMETERS THAT CHARACTERIZE THE DISTRIBUTIONS FOR AMOUNTS OF EXTREME VALUES IN COORDINATE DISTRIBUTIONS $\delta(m \times n)$ OF LASER IMAGES FOR HUMAN BILE

$\delta(m \times n)$	$\delta(m \times n) = 0$		$\delta(m \times n) = \pi$	
	Healthy (21 patients)	Diabetes (19 patients)	Healthy (21 patients)	Diabetes (19 patients)
M_1^{δ}	0.51±0.063	0.54±0.067	0.22±0.025	0.35±0.042
M_2^{δ}	0.13±0.018	0.08±0.011	0.25±0.031	0.14±0.017
M_3^{δ}	0.26±0.033	0.19±0.022	0.79±0.086	2.18±0.25
M_4^{δ}	0.48±0.054	0.55±0.068	0.83±0.098	3.11±0.42
R_4^{δ}	0.14±0.016	0.12±0.015	0.56±0.069	2.21±0.31
S^{δ}	0.24±0.015	0.21±0.013	0.17±0.021	0.08±0.012
F^{δ}	2.42±0.12	2.49±0.11	2.58±0.15	Statistical
D^{δ}	0.21±0.028	0.24±0.027	0.34±0.042	0.18±0.023

- correlation area S^{δ} for the autocorrelation dependence $K_{\max}(\Delta x)$ of the distribution for the amount of extreme phase shifts in a laser image inherent to joint bile of a patient with osteoarthritis is 2.55 times less than that parameter determined for a healthy patient;
- distributions $N_{\max}(x)$ for the phase maps describing bile for healthy and sick patients are, respectively, fractal and statistical;
- dispersion D^{δ} of the dependences $\log J(N_{\max}) - \log d^{-1}$ in the case of pathological changes in the polycrystalline structure of bile is 1.75 times decreased.

VI. CONCLUSIONS

Thus, one can draw the following conclusions:

➤ Human bile, independently of their physiological state, contains phase-modulating optically anisotropic network of biological crystals.

➤ Ascertained and grounded is a set of criteria for phase diagnostics of inflammatory processes (diabetes, cholecystitis) as being based on statistical (statistical moments of the first to fourth orders), correlation (statistical moments for the Gramm-Charlie expansion, correlation area) and fractal (fractal dimensionality and dispersion for the distribution of extrema in log – log dependences of power spectra) analyses of phase distributions in laser images of human bile.

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